

**Monoamine Oxidase A  
ELISA Kit Format I  
96T**

General reference manual. Use the protocol shipped with the kit for your experiments.

FIVEphoton  
Biochemicals

**For research use only.  
Not for diagnostics.**

Reference manual for parts

hMAO-A-ELISA1

mMAO-A-ELISA1

rMAO-A-ELISA1

pMAO-A-ELISA1

FIVEphoton Biochemicals  
4907 Morena Blvd, Ste 1403  
San Diego, CA 92117  
Tel: 858 395-4026  
Fax: 858 345-5291  
Website: [www.fivephoton.com](http://www.fivephoton.com)  
[customersupport@fivephoton.com](mailto:customersupport@fivephoton.com)

**Store:** 2-8°C. Expiration: 6 months after arrival.

**Safety:** Stop solution contains acid. Avoid contact and inhalation. Wear eye protection.

# Monoamine Oxidase ELISA A Kit– Format 1

## Part Nos. h,r,m,p MAO-A ELISA1

**For research only. Not for diagnostic applications.**

Storage: 2-8°C, expires 6 months after arrival

Safety: Stop solution contains acid. Avoid eye and skin contact

Standard Peptides:

Human: 128ng/ml

Mouse: 128ng/ml

Rat: 128ng/ml

Porcine: 128ng/ml

Assay Ranges

Human: 500 pg/ml - 100 ng/ml

Mouse: 500 pg/ml – 100 ng/ml

Rat: 500 pg/ml – 100 ng/ml

Porcine: 500 pg/ml – 100 ng/ml

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**Overview:** Please read this protocol completely prior to using the product. The kit measures Monoamine Oxidase A (MAO-A) in sandwich ELISA format. The ELISA plate is provided pre-coated with an anti-MAO-A monoclonal antibody. The experimenter mixes the samples with a polyclonal detection anti-MAO-A antibody (that is provided biotin labeled) with the provided streptavidin-HRP solution *prior* to the addition of the solutions to the ELISA plate. The sample-antibody-biotin-streptavidin-HRP mixture is then added to the ELISA plate, which is incubated for 60 min at 37°C. Unbound biotin-linked antibody is washed away and color reagents are added that change the color of the solution to blue. The blue solution then turns yellow after addition of the stop solution. The absorbance of the resulting yellow solution at 450nm corresponds to the concentration of Monoamine Oxidase A in the samples.

**Sample Preparation - The below is provided as a generic overview. Please review the literature to identify the optimal method to isolate your target protein**

1. **Serum:** Coagulate at room temperature for 10-20 min. Centrifuge for 20 min at 2000-3000 rpm. Collect supernatant for assay. If precipitation appears, centrifuge again. Assay the supernatant fraction.
2. **Plasma:** Use suitable EDTA or heparin as an anticoagulant. Mix for 10-20 min using a stir bar. Centrifuge for 20 min at 2000-3000 rpm. Collect supernatant. If precipitation appears, centrifuge again and collect the supernatant.
3. **Urine:** Collect in a sterile container. Centrifuge 20-min at 2000-3000 rpm. Collect supernatant, If precipitation appears, centrifuge again. Collect supernatant for assay.
4. **Cell culture supernatant:** Detection of extracellular components: Centrifuge culture media for 20 min at 2000-3000 rpm. Collect supernatant for assay.
5. **Membrane fraction:** Lyse cells with a non-denaturing detergent buffer (such as FIVEphoton Biochemicals part no. ELSP-1). Sediment debris and assay the clarified supernatant fraction.
5. Samples can be store at -80°C. Avoid repeated freeze-thaw cycles. You may aliquot samples for later ELISA assays.
6. **Avoid denaturing cell lysis buffers that contain SDS such as RIPA buffer.**

7. Samples can be diluted in a sample dilution solution with the formula: 1% BSA in PBS pH 7.4. A 1:5 dilution is recommended. i.e. 10 µl sample, 40 µl sample dilution solution.

**Table 1. Materials supplied. Store all materials at 4°C**

1	Standard peptide: 128 ng/ml	0.5ml	7	Chromogen Solution A	6 ml
2	Standard diluents (1% BSA in PBS pH 7.4)	3 ml	8	Chromogen Solution B	6 ml
3	Anti-MAO-A Coated ELISA plate (96T)	12 wellx8 strips	9	Stop Solution	6 ml
4	Streptavidin-HRP	6 ml	10	Instruction Manual	1
5	30xwash solution	20 ml	11	Closure Membrane	2
6	MAO-A antibody labeled with biotin (detection antibody)	1ml	12	Sealed bags	1

**Materials required but not supplied**

1. Sample dilution buffer (1% BSA in PBS pH 7.4)
2. 37°C incubator
3. Standard absorbance plate reader
4. Precision pipettes and disposable pipette tips
5. De-ionized water
6. Disposable tubes for sample dilution
7. Absorbent paper

**Important notes and preparation for the assay**

1. Make a solution that is 1% BSA in PBS pH 7.4 for dilution of samples (i.e."sample dilution solution").
2. Equilibrate the ELISA plate strips to room temperature for at least 30 minutes. The plate strips should be kept in a zip lock bag to prevent drying of the wells. Store unused sections of the ELISA plate in a sealed bag at 4°C.
3. Use new pipettes tips at each step to prevent contamination.
4. Do not use reagents from other ELISA kits..
5. Substrate B is light sensitive. Limit light exposure.
6. **Manual Wash Method:** Gently aspirate off the liquid in each well. Turn over the plate and pat-dry on absorbent paper. Add 100µl wash solution and repeat 5 times. Therefore 600ul wash solution is needed per well. An automatic washer can also be employed to wash the ELISA wells.

**Assay procedures**

Blanks, standard and sample preparation. Prepare the blanks, standards and samples in a separate 96-well dish and transfer simultaneously to the ELISA plate. Note that the sample – detection antibody – streptavidin - HRP complex should be prepared by the experimenter prior to addition of the mixture to the ELISA plate wells: Do not prepare samples in the ELISA plate.

**Assay procedure**

1. Standard dilutions guide. The following table is provided for Human Monoamine Oxidase A ELISA. Calculate similar dilutions for other species. Use Table 2 as a guide for dilutions of standards in the provided standard diluent

solution. Make dilutions in separate tubes or multi-well dishes, not in the ELISA dish. Use 2 replicates for the blank, standard and sample wells.

**Table 2. Standard Dilutions for Human Monoamine Oxidase A ELISA. Use a similar dilution scheme for other species.**

Standard Concentration	Standard Number	Dilution Instructions
64 ng/ml	7	120µl original standard + 120µl standard diluents solution, mix
32 ng/ml	6	120µl standard No. 7 + 120µl standard diluents solution, mix
16 ng/ml	5	120µl standard No. 6 + 120µl standard diluents solution, mix
8 ng/ml	4	120µl standard No. 5 + 120µl standard diluents solution, mix
4 ng/ml	3	120µl standard No. 4 + 120µl standard diluents solution, mix
2 ng/ml	2	120µl standard No. 3 + 120µl standard diluents solution, mix
1 ng/ml	1	120µl standard No. 2 + 120µl standard diluents solution, mix

2. Blank, standard and sample preparation prior to addition to the ELISA wells:
  - a) Blank wells: Set aside 2 blank wells. Only add chromogen solutions A and B and then stop solution later during the procedural steps while developing the ELISA.
  - b) Standard solution preparation: Add 50µl standard solution (that was previously diluted) and 50µl streptavidin-HRP solution to each standard. Detection antibody labeled with biotin has been added in advance to the standards, therefore anti-MAO-A detection antibody is not added to prepare the standards solutions.
  - c) Sample wells: Dilute 10µl sample in 30µl sample diluent solution (1% BSA in PBS pH 7.4 – sample diluent is made by the experimenter), then add 10µl anti-MAO-A antibody labeled with biotin (detection antibody) and 50µl streptavidin-HRP. Mix gently; do not vortex.
  - d) Transfer the standard and sample solutions that you have prepared simultaneously to the ELISA dish. Seal the wells with a ziplock bag and rotate the plate gently for 60 minutes at 37°C.
3. Preparation of wash solution: Dilute the 30X wash solution to 1X with dH<sub>2</sub>O. Prepare 600µl 1X wash solution per well.
4. Wash: Carefully remove the seal plate membrane or zip lock bag that protects the wells from drying in a manner that does not cross-contaminate the wells with liquid. Aspirate off the liquid. Fill each well with 100µl 1X wash solution. After 1 minute, aspirate off the wash solution. Then repeat the wash procedure five times for 30 seconds, each time. Quickly blot dry the plate.
5. Color development: *Chromogen Solution B is light sensitive; protect from light.* Simultaneously transfer 50µl of chromogen solution A to each well and then simultaneously transfer 50µl of chromogen solution B to each well. *Protect the plate from light.* Incubate the plate protected from light for 10 minutes at 37°C.
6. Stop solution: Add 50µl Stop Solution to each well to stop the reaction: Set the blank well as zero, and measure the absorbance (OD) of each well at 450nm within 15 minutes after adding the stop solution.

### Data Analysis

1. Compile a standard curve using the blank standard solutions and the corresponding OD values. You may wish

to calculate a linear regression equation from the standard curve to determine the concentration of your samples. Take into account dilution factors in your calculation of UB concentration. Alternative methods of data analysis to calculate concentrations of your samples can also be employed.

## **Flow chart of the procedures**

Separately prepare standards, blank and samples with detection antibody and HRP-streptavidin as required



Incubate with the ELISA plate for 1 hr at 37°C.



Wash the plate five times.



Add Chromogen solutions A and B. Incubate for 10 min at 37°C for color development.



Add stop solution



**Measure OD value at 450nm within 10 min**